# UTSouthwestern Medical Center

## Department of Surgery

### BACKGROUND



Metastasis and resistance to chemotherapy remain the major challenges in the treatment of pancreatic ductal adenocarcinoma (PDAC) patients. Tumorintrinsic and suppressive immune mechanisms contribute to chemotherapy failure. Strategies that enhance the efficacy of standard therapies are likely the most efficient route to improve overall survival in this disease.

Axl is a member of the TAM (Tyro3, Axl, MerTK) receptor tyrosine kinase family and participates in various cellular processes, including cell survival, proliferation, migration and epithelial plasticity. Growth arrest specific factor 6 (Gas6) is the ligand for Axl. Once Gas6 binds to Axl, it induces the dimerization of the Gas6/Axl binding complex. Ligand-induced activation of Axl occurs only after the Gla domain of Gas6 binds to phosphatidylserine. The Gas6-Axl pathway is estimated to be active in 70% of human pancreatic cancers where it correlates with an adverse clinical prognosis. Axl expression and activity are also associated with metastasis, therapy resistance, and immunosuppression in various types of cancers. Here we demonstrate the function of AxI in pancreatic cancer cellular plasticity and microenvironment, highlighting the importance of AxI on PDAC progression, metastases and

## sensitivity to gemcitabine in *KIC*



were determined by detecting recombined *Ink4a/Arf* in liver tissues. Data are displayed as mean ± SD. \*P < 0.05; by t-test. **B)** KIC and Axl<sup>LacZ/LacZ</sup> KIC mice were enrolled in a survival study and randomized to vehicle and gemcitabine (Gem, 25) mg/kg ip twice a week). Therapy was initiated on day 45 and maintained until sacrifice.

### II. AxI-deficient KIC displays a more differentiated histology and has a more inflammatory and active immune microenvironment



## Axl is critical for pancreatic cancer cellular plasticity and metastasis Wenting Du<sup>1</sup>, Huocong Huang<sup>1</sup>, Jason Toombs<sup>1</sup>, Natalie Burton<sup>1</sup>, Zhaoning Wang<sup>2</sup>, Yuqing Zhang<sup>1</sup>, Rolf A. Brekken<sup>1</sup>

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A) Single cell RNA sequencing was performed with tumors harvested from KIC or KPfC (Kras<sup>LSL-G12D</sup>, P53<sup>lox/lox</sup>, PDX<sup>Cre/+</sup>) mice. tSNE plots of the KIC and KPfC tumors and violin plots of Axl expression for each cell population are shown. Axl expression is highlighted in red in the second tSNE plot for each mouse model. B) Correlation plots with AxI and EMT genes in KIC and KPfC models were generated by analyzing the EMT gene expression within the cancer cell clusters from each model. A positive correlation is highlighted in red and negative correlation is highlighted in blue. C) Tumor tissues from Axl<sup>LacZ/+</sup> KIC mice were evaluated for co-localization of β-galactosidase and tumor cell marker Sox9, epithelial marker E-Cadherin and mesenchymal marker vimentin.  $\beta$ -galactosidase staining is used to identify the expression of AxI.





A) AxI was ablated out using CRISPR-Cas9 in KPfC tumor cells. Cell proliferation was investigated in WT and AxI KC KPfC cells using MTS assay. B) Cell migration was investigated in WT and AxI KO KPfC cells by a "scratch" assay. Monolayers of the indicated cells were wounded with a pipet tip. The cells were incubated in 1% serum media. Wound closure was monitored at 12, 24, and 36 hours and is reported as % wound closure. \*\*\*P < 0.001; \*\*\*\*P < 0.0001; by ttest. C) Colony formation for WT and AxI KO KPfC cells is shown as mean + SD colonies/hpf . \*P < 0.05; by t-test. D) Cell lysates from WT and AxI KO KPfC cells were evaluated for indicated targets by western blot. E) Representative pictures of WT and AxI KO KPfC cells grown in 30% matrigel+70% collagen for 4 days. F) WT and AxILacZ/LacZ C57BL/6 orthotopically injected with WT and AxI KO KPfC cells were enrolled in a survival study. Groups were as follows: WT KPfC cells in WT C57BL/6 mice; WT KPfC cells in Axl<sup>LacZ/LacZ</sup> C57BL/6 mice; Axl KO KPfC cells in WT C57BL/6 mice; Axl KO KPfC cells in Ax/LacZ/LacZ C57BL/6 mice. G) Number of gross liver metastases and metastatic incidence of the above survival study are shown. \*\*\*\*P < 0.0001 vs WT KPfC cells in WT C57BL/6 mice; ## P < 0.01 vs WT KPfC cells in Axl<sup>LacZ/LacZ</sup> C57BL/6 mice; by ANOVA.

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Ludwig and Du, et al. Cancer research, 2018 A) KIC mice were enrolled in a survival study and randomized to vehicle (Control), BGB324 (50 mg/kg PO BID), gemcitabine (25 mg/kg ip twice a week), and gemcitabine + BGB324. Therapy was initiated on day 49 and maintained until sacrifice. B) Panc 265 was orthotopically implanted into nu/nu athymic mice. After tumor establishment, mice were treated as in (A) (n = 6-7/group). Total gross metastasis was determined by evaluation of liver, diaphragm, GI lymph nodes, and lung at the time of sacrifice. BGB324 alone or in combination with gemcitabine significantly reduced the rate of metastasis (p=0.001 vs control, Fisher's exact test). C) and D) Tumor tissues from KIC mice above were evaluated for indicated targets for epithelial (E-Cadherin) to mesenchymal (Vimentin) transition or immunosuppressive (Arginase 1) macrophages (F4/80). Images were analyzed using Elements software; quantification of % area fraction is shown. Data are displayed as mean ± SD and represent 5 images per tumor with 4 animals per group analyzed. \*\*P < 0.01; \*\*\*P < 0.005; \*\*\*\*P < 0.001; by ANOVA. E) Flow cytometry of PDAC tumors treated with vehicle (Control) or BGB324 (50 mg/kg, PO, BID) for two weeks. BGB324 reduced monocytic MDSCs (M-MDSCs, CD11b+ Ly6G- Ly6C+), PD-L1+ M-MDSCs, tumor associated macrophages (CD11b+ Ly6G- Ly6C- F4/80+ CD11c+ MHCII+) and Arginase1+ tumor associated macrophages. \*\*, P < 0.01; \*\*\*, P < 0.005 by t-test.

Pancreatic ductal adenocarcinoma, a leading cause of cancer-related death in the US, has a high metastatic rate that is associated with persistent immune suppression. Axl, a member of the TAM (Tyro3, Axl, MerTK) receptor tyrosine kinase family, has been identified as a critical factor that drives metastasis and immune suppression in many cancer types. Using genetically engineered mouse models, we demonstrate that:

- to chemotherapy (gemcitabine) compared to AxI WT PDAC mice.
- microenvironment
- Axl is expressed highly in tumor cells that have a mesenchymal-like phenotype.

To study the contribution of AxI positive mesenchymal tumor cells, AxI was knocked out using CRISPR-Cas9 in primary tumor cells isolated from KPfC tumors. Characterizing these cells shows that: • Axl-deficient PDAC cells displayed a more differentiated phenotype in vitro compared to Axl WT parental cells.

Axl expression on tumor cells is critical for PDAC progression and metastasis.

As a potential therapeutic target, pharmacological inhibition of AxI by BGB324 significantly reduces tumor progression and metastasis and improves response to chemotherapy in multiple preclinical PDAC models. As a result, clinical testing of BGB324 (Bemcentinib) in conjunction with standard chemotherapies in pancreatic cancer patients has been initiated (NCT03649321). Bemcentinib, as well as other strategies of Axl inhibition, is also being investigated as a single agent or in combination with targeted-, chemo-, and immunotherapy in different types of cancer patients.

### CONCLUSION

• Axl-deficient PDAC bearing mice have longer median survival, smaller tumors, fewer metastases and higher sensitivity

• Axl-deficient PDAC displays a more differentiated histology and has a more inflammatory and active immune